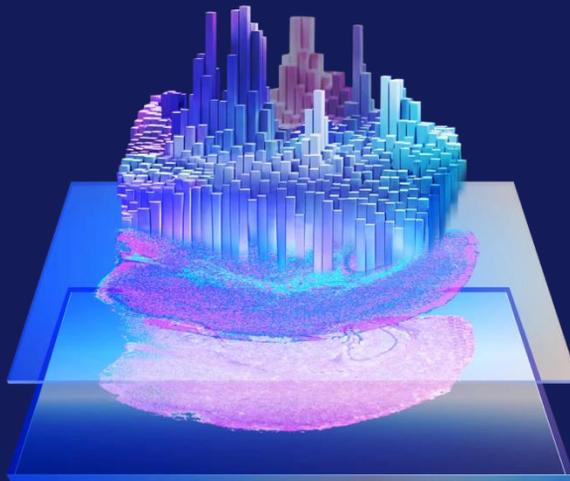
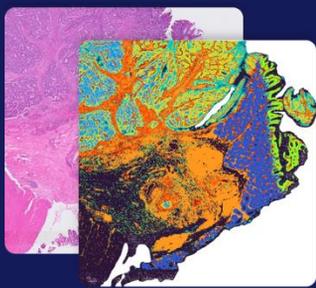


## 10x Genomics Visium HD Training Three-Day Hands-On Course on Spatial Transcriptomics

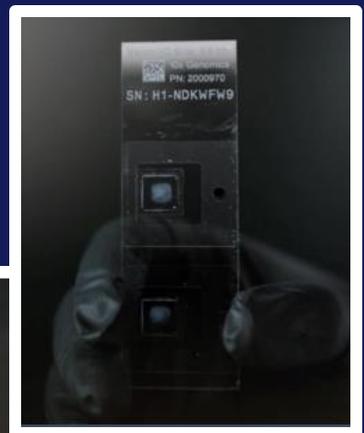
**MRC WIMM Advanced Single Cell Omics Facility (WASCOF),  
University of Oxford**



**Whole transcriptome discovery in the tissue context!**

### Learning Objectives

- FFPE Samples Requirement
- Tissue Sectioning and H&E (IF)
- Mastering the Visium CytAssist
- Visium Library Construction
- Quality Control
- Sequencing guidelines
- Loupe Browser



**Dates: Monday 23<sup>rd</sup> – Wednesday 25<sup>th</sup> September 2024**  
**Venue: MRC Weatherall Institute of Molecular Medicine, John Radcliffe Hospital, Headington, Oxford OX3 9DS**

**Register Here:**

<https://forms.gle/MkZ2VTvHVg8nDBch8>

or email [Maria.Greco@imm.ox.ac.uk](mailto:Maria.Greco@imm.ox.ac.uk)

# Course Program

Presenters: Dr Claudia Tamburini (FAS 10x Genomics); Dr Maria Greco (MRC WIMM); Dr Esther Bridges (MRC WIMM); Dr Paola Vargas Gutierrez (MRC WIMM); Dr Lai Cheng (MRC WIMM)

## Monday 23<sup>rd</sup> September

Time	Topic
10.00	<b>Talk: Visium HD_Assay overview and sample preparation. WIMM Conference Room</b>
11.15	Coffee break
11.45	Practical Session: Sectioning and Mounting. <i>Training Lab</i>
13.15	Lunch Break
14.00	Deparaffinization and H&E Staining, Tissue Imaging & Decrosslinking. <i>Training Lab</i>
16.30	Probe Incubation (O.N.). <i>Training Lab</i>
16.45	Coffee Break Q&A: Conclusions, participant Q&A and wrap up of Day 1

## Tuesday 24<sup>th</sup> September

9.30	Post Hybridization Wash & Probe Ligation. <i>Training Lab</i>
11.10	Visium HD Slide Preparation. <i>Training Lab</i>
11.30	Visium CytAssist Session for Probe release and Capture. <i>Training Lab</i>
13.30	Lunch Break
14.00	Pre-Amplification and SPRI-select. <i>Training Lab</i>
15.00	<b>Talk: Visium HD_Assay workflow and technical considerations. WIMM Conference Room</b>
16.30	Coffee Break Q&A: Conclusions, participant Q&A and wrap up of Day 2

## Wednesday 25<sup>th</sup> September

9.30	Cycle number determination – qPCR. <i>Training Lab</i>
10.10	qPCR result explained & consideration. <i>Training Lab</i>
10.45	Sample Index PCR. <i>Training Lab</i>
11.00	Coffee Break
11.30	Post-sample Index PCR Cleanup - SPRI-select. <i>Training Lab</i>
12.00	Post-Library construction QC & Bioanalyzer results explained. <i>Training Lab</i>
13.00	Lunch Break
14.00	<b>Data Overview_ Loupe Browser demo. WIMM Conference Room</b>
16.00	Coffee Break Q&A: Conclusions, participant Q&A and course wrap up.