Downloading published fastq data from GEO

This guide will show you how to download fastq format data from published papers.

Look in the paper for the GEO accession number and then go to the GEO website: <u>http://www.ncbi.nlm.nih.gov/geo/</u>

Enter the GEO accession code for that data you want to download (e.g. **GSE37757**) and click Search

			<u>S</u>	ign in to NCBI		
GEO Home Documentation - Query	v & Browse 👻 Email GEO					
Gene Expression Om	nibus		Gene Expre			
sequence-based data are accepted. Tools are gene expression profiles.	provided to help users query and download experiments and curated	1	Keeward or GEO Accession	Search		
			Reyword of GEO Accession	ocuren		
Getting Started	Tools	Browse Conten	it			
Overview	Search for Studies at GEO DataSets	Repository Browser	Repository Browser			
FAQ	Search for Gene Expression at GEO Profiles	DataSets:	3413			
About GEO DataSets	Search GEO Documentation	Series: 🔝	40497			
About GEO Profiles	Analyze a Study with GEO2R	Platforms:	11892			
About GEO2R Analysis	GEO BLAST	Samples:	974975			
How to Construct a Query	Programmatic Access					
How to Download Data	FTP Site					
Information for Submitters						
Login to Submit	Submission Guidelines	MIAME Standards				
	Update Guidelines	Citing and Linking to	Citing and Linking to GEO			
		Guidelines for Revie	wers			

You will then see a page for that **Series**:

S NCBI	
	GEO Publications EAO MIAME Email GEO
NCDI > GEO > ACCES	
GEO help: Mouse ove	er screep elements for information.
Scope: Self \$	Eormat: HTML + Amount: Quick + GEO accession: GSE37757 GO
Series GSE3775	Query DataSets for GSE37757
Status	Public on Jul 01, 2012
Title	Unraveling cell type-specific and reprogrammable human replication origin signatures associated with G-quadruplex consensus motifs
Organism	Homo sapiens
Experiment type	Genome binding/occupancy profiling by high throughput sequencing
Summary	Short nascent strands purification coupled to next-generation sequencing allowed us to identify replication origins on human genome in an extensive way, by mapping replication origins in 4 different cell types, IMR-90 fibroblasts, hESC H9 cells, iPSC Th CI-4 cells and HeLa cells. We demonstrated the existence of a cell type-specific reprogrammable signature of the cell identity revealed by specific efficiencies of conserved origin positions and not by the selection of cell-type specific subsets of origins.
Overall design	4 different cell types were analyzed. For each cell types, 2 different biological replicates of short nascent strands at replication origins were purified. Each SNS sample was sequencing at least one time.

Scroll down that page to find the 'Samples' section and click 'More' link if necessary to see all the samples in the entry.

		Gene Expression Omnibus	
IME SEARCH SITE MA	2	GEO Publications FAQ	MIAME Email GE
CBI > GEO > Acces	sion Display 🛛		Not logged in Logir
GEO help: Mouse over	screen elements for information.		
Scope: Self \$	Format: HTML + Amount:	Ouick + GEO accession: GSE37757	GO
Series GSE3775	7	Query DataSets for GSE3	37757
Status	Public on Jul 01, 2012		
Title	Unraveling cell type-specific a signatures associated with G-qua	nd reprogrammable human replication druplex consensus motifs	origin
Organism	Homo sapiens		
Experiment type	Genome binding/occupancy profi	ling by high throughput sequencing	
Summary	allowed us to identify replication by mapping replication origins in H9 cells, iPSC Th Cl-4 cells and H We demonstrated the existence of the cell identity revealed by s and not by the selection of cell-ty	A different cell types, IMR-90 fibroblasts, eLa cells. of a cell type-specific reprogrammable sig specific efficiencies of conserved origin po ype specific subsets of origins.	e way, hESC nature sitions
Overall design	4 different cell types were anal replicates of short nascent stran sample was sequencing at least of	yzed. For each cell types, 2 different bic ds at replication origins were purified. Eac one time.	ological h SNS
Contributor(s)	Besnard E, Babled A, Lapasset L, Lemaitre J	Milhavet O, Parrinello H, Dantec C, Marin	з,
Citation(s)	Besnard E, Babled A, Lapasset L, and reprogrammable human rep quadruplex consensus motifs. <i>Na</i> PMID: 22751019	Milhavet O et al. Unraveling cell type-spec lication origin signatures associated with O It Struct Mol Biol 2012 Aug;19(8):837-44	cific 5-
Submission date	May 04, 2012		
Last update date	Jul 01, 2013		
Contact name	Emilie Besnard		
Organization name	Institute of Functional Genomics		
Lab	Genome Plasticity and Aging		
Street address	141 rue de la Cardonille		
City ZID/Destal and	MONTPELLIER		
Country	France		
country .	- rande		
Platforms (1)	GPL9115 Illumina Genome Anal	yzer II (Homo sapiens)	
Samples (4)	GSM927235 IMR-90		
≝ More	GSM927236 hESC H9		
	GSM927237 iPSC Thomson clon	e 4	
Relations			
BioProject	PRJNA163241		
SRA	SRP012667		
Download family		Format	
SUFT formatted far	niy nie(s)	SOFT 😢	
1471-1140 A	5		

Click on one the sample links eg 'GSM927238'

You will now see an entry for that sample

Scope: Self Format: HTML Amount: Quick GED access Sample GSM927238 Query Status Public on Jul 01, 2012 Title HeLa Sample type SRA Source name cervical cancer cells Organism Homo sapiens Characteristics cell line: HeLa Treatment protocol No specific treatment Growth protocol HeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal B % L-Glutamine (200 mM), 1 % penicillin and strepton Extracted molecule genomic DNA Extraction protocol We purified short nascent strands at replication origin realized isolation of Short Nascent Strands at replication origin in realized isolation of Short Nascent Strands at replication origin synchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was read DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing purified background DNA to the sequencing purified background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing purified and sonit to background DNA to the sequencing purified and sonit to short nascent strands (SNS) at replic exonuclease	
Sample GSM927238 Query Status Public on Jul 01, 2012 Title HeLa Sample type SRA Source name cervical cancer cells Organism Homo sapiens Characteristics cell line: HeLa Treatment protocol HeLa cells were maintained in Dulbecco's Modifie Growth protocol HeLa cells were maintained in Dulbecco's Modifie Status % L-Glutamine (200 mM), 1 % penicillin and strepton Extracted molecule genomic DNA Extraction protocol Extraction protocol We purified short nascent strands at replication origin realized isolation of Short Nascent Strands at replication was us millions asynchronous numan cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was read DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of background around the replication origins to be able to compare the replication origins between same amount of background DNA to the sequencing proto be able to compare the replication origins between same amount of background DNA to the sequencing proto background DNA to the sequencing proto background DNA to the sequencing protoxis background DNA to the sequencing protoxis background prote tor	ssion: GSM927238 GO
Sample GSM927238QueryStatusPublic on Jul 01, 2012TitleHeLaSample typeSRASource namecervical cancer cellsOrganismHomo sapiensCharacteristicscell line: HeLaTreatment protocolNo specific treatmentGrowth protocolHeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal Bu % L-Glutamine (200 mM), 1 % penicillin and streptonExtracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin realized isolation of Short Nascent Strands at replication genomic Cadoret et al. PNAS 2008). For each cell type, wa biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was rea DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of back the basis of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing pLibrary strategyOTHER Library source genomic Library selectionLibrary selectionother Instrument modelIllumina Genome Analyzer IIDescriptionenrichment of short nascent strands (SNS) at replic exonucleaseData processingThe next generation sequencing of purified and somi	
Status Public on Jul 01, 2012 Title HeLa Sample type SRA Source name cervical cancer cells Organism Homo sapiens Characteristics cell line: HeLa Treatment protocol No specific treatment Growth protocol HeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal Bink % L-Glutamine (200 mM), 1 % penicillin and strepton Extracted molecule genomic DNA Extraction protocol We purified short nascent strands at replication origin: realized isolation of Short Nascent Strands at replication origin: realized isolation of Short Nascent Strands at replication origin: realized isolation SNS preparation was tested, as previ al. PNAS 2008). For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambdid for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was rea DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of back the basis of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing p Library strategy OTHER Library selection other Instrument model Illumina Genome Analyzer II Description enrichment o	/ DataSets for GSM927238
TitleHeLaSample typeSRASource namecervical cancer cellsOrganismHomo sapiensCharacteristicscell line: HeLaTreatment protocolNo specific treatmentGrowth protocolHeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal Bi % L-Glutamine (200 mM), 1 % penicillin and streptonExtracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNA/ ragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was rea DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of back ground around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing pLibrary strategyOTHER Library source genomicLibrary selectionother Instrument modelIllumina Genome Analyzer IIDescription exonucleaseData processingThe next generation sequencing of purified and soni	
Sample typeSRASource namecervical cancer cellsOrganismHomo sapiensCharacteristicscell line: HeLaTreatment protocolNo specific treatmentGrowth protocolHeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal Br % L-Glutamine (200 mM), 1 % penicillin and streptonExtracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was ree DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of back the basis of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing pLibrary strategyOTHER Library source genomicLibrary selectionother Instrument modelIllumina Genome Analyzer IIDescription exonucleaseData processingThe next generation sequencing of purified and soni	
Source namecervical cancer cellsOrganismHomo sapiensCharacteristicscell line: HeLaTreatment protocolNo specific treatmentGrowth protocolHeLa cells were maintained in Dulbecco's ModifieInvitrogen) containing 10 % heat-inactivated Fetal Bi % L-Glutamine (200 mM), 1 % penicillin and streptonExtracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin: realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was read DNA labeling system. In order to be reproducible, we same purified solation origins between same amount of background DNA to the sequencing purified the basis of background DNA to the sequencing purified Library selectionLibrary selectionother Illumina Genome Analyzer IIDescriptionenrichment of short nascent strands (SNS) at replic exonucleaseData processingThe next generation sequencing of purified and sonities	
OrganismHomo sapiensCharacteristicscell line: HeLaTreatment protocolNo specific treatmentGrowth protocolHeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal Bi % L-Glutamine (200 mM), 1 % penicillin and streptonExtracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was read DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing pLibrary strategyOTHER Library selection otherLibrary selectionother Illumina Genome Analyzer IIDescriptionenrichment of short nascent strands (SNS) at replic exonucleaseData processingThe next generation sequencing of purified and sonit	
Characteristicscell line: HeLaTreatment protocolNo specific treatmentGrowth protocolHeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal Bi % L-Glutamine (200 mM), 1 % penicillin and streptonExtracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin: realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was ree DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of back the basis of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing pLibrary strategyOTHER Library selection otherLibrary selectionotherInstrument modelIllumina Genome Analyzer IIDescriptionenrichment of short nascent strands (SNS) at replic exonucleaseData processingThe next generation sequencing of purified and soni	
Treatment protocol No specific treatmentGrowth protocolHeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal Ba % L-Glutamine (200 mM), 1 % penicillin and streptonExtracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was rea DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of back the basis of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing pLibrary strategyOTHER Ilibrary selection otherLibrary selectionIllumina Genome Analyzer IIDescriptionenrichment of short nascent strands (SNS) at replic exonucleaseData processingThe next generation sequencing of purified and soni	
Growth protocolHeLa cells were maintained in Dulbecco's Modifie Invitrogen) containing 10 % heat-inactivated Fetal Br % L-Glutamine (200 mM), 1 % penicillin and streptonExtracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was rea DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of back the basis of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing pLibrary strategyOTHER Ilibrary selection otherDescriptionenrichment of short nascent strands (SNS) at replic exonucleaseData processingThe next generation sequencing of purified and soni	
Extracted molecule genomic DNAExtraction protocolWe purified short nascent strands at replication origin realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAcol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd for each purified SNS preparation was tested, as previ al. PNAS 2008). Synthesis of second strands was rea DNA labeling system. In order to be reproducible, we sample, this synthesis with the same amount of back the basis of background around the replication origin to be able to compare the replication origins between same amount of background DNA to the sequencing pLibrary strategyOTHER Library selection Illumina Genome Analyzer IIDescriptionenrichment of short nascent strands (SNS) at replic exonucleaseData processingThe next generation sequencing of purified and soni	d Eagle Medium (DMEM, ovine Serum (FBS, PAA), 1 nycin (all from Invitrogen).
Extraction protocolWe purified short nascent strands at replication origin: realized isolation of Short Nascent Strands at re asynchronous human cells by adapting the prot (Cadoret et al. PNAS 2008). For each cell type, we biological replicates. For each sample, DNAzol was us millions asynchronous cells. Then DNA fragments of sucrose gradient separation. After two runs of lambd 	
Library strategy OTHER Library source genomic Library selection other Instrument model Illumina Genome Analyzer II Description enrichment of short nascent strands (SNS) at replice exonuclease Data processing The next generation sequencing of purified and sonice	s in different cell types. We plication origins (SNS) of tocol previously described e purified the SNS of two sed to extract DNA of 100 1-2 kb were isolated after da exonuclease, the quality iously described (Cadoret et alized with the kit bioprime realized, for each biological kground DNA calculated on a the Myc locus. In order different cells, we gave the platform.
Library source genomic Library selection other Instrument model Illumina Genome Analyzer II Description enrichment of short nascent strands (SNS) at replice exonuclease Data processing The next generation sequencing of purified and soni	
Library selection other Instrument model Illumina Genome Analyzer II Description enrichment of short nascent strands (SNS) at replice Data processing The next generation sequencing of purified and sonic	
Instrument model Illumina Genome Analyzer II Description enrichment of short nascent strands (SNS) at replic exonuclease Data processing The next generation sequencing of purified and soni	
Description enrichment of short nascent strands (SNS) at replic exonuclease Data processing The next generation sequencing of purified and soni	
Data processing The next generation sequencing of purified and soni	cation origins with lambda
the Montpellier GenomiX (MGX) facility in Mont Illumina's sequencing by synthesis technology. The clusterized and then hybridized with sequencing prime the mode by clusterized and the sequencing prime	icated SNS was realized in pellier, France using the resequencing library was hers. The sequencing of 36

Scroll down until you see the 'Relations' section and click on the SRA link:

Series (1)	GSE37757 Unraveling cell type–specific and reprogrammable human replication origin signatures associated with G-quadruplex consensus motifs	
Relations		
SRA	SRX145994	
BioSample	SAMN00990948	

Next you will see a 'SRA' page. Note the number of runs that make up this entry (in this case, 4) and note the SRR numbers for each:

					Auvaliceu
Disp	olay Settings	s: 🕑 Full			<u>Send to:</u> ⊘
SRX	(145994: GSI	M927238: HeLa;	Homo sapiens;	OTHER	
ILL	LUMINA (Illui	mina Genome A	nalyzer II) runs: 2	52.4M spots,	9.1G bases, 5.6Gb downloads
Acce	ession: SRX	145994			
Expe	eriment des	ign: n/a			
sub	mission: SF	A052331 by GE	0		
Stud	dy summary	: GSE37757: Ge	enome-wide mapp	ing of replicat	tion origins reveals cell-type specific and reprogrammable replication origin signatures in
uma	an ceils (SRI	P012667) • <u>Stud</u> RS312185 (mor	y • <u>All experiment</u>	ts (more)	
sam					
sam _ibra	arv: GSM92	7238: HeLa (mo	ent		
ibra Ibra Iatf	ary: GSM92 form: Illumin	7238: HeLa (mo a (more)	pre)		
ibra libra latf Spot	ary: GSM92 form: Illumin t descriptor:	7238: HeLa (mo a (more)	e)		
ibra latf Spot	form: Illumin t descriptor: forward	7238: HeLa (mo a (more)	re)		
ibra latf Spot	form: Illumin t descriptor: forward	7238: HeLa (mo a (more))		
ibra Platf Spot	ary: GSM92 form: Illumin t descriptor: forward eriment attri	7238: HeLa (more)) re)		
ibra Platf Spot	eriment attri	2238: HeLa (mo a (more) butes: m: GSM927238	2002 (re)		
ibra Platf Spot 1 Expe GL Tota	ary: GSM92 form: Illumin t descriptor: forward eriment attri EO Accessio al: 4 runs, 25	(more) (more) (butes: nr: GSM927238 2.4M spots, 9.10	G bases, <u>5.6Gb</u> (2 🔇	
ibra Platf Spot	ary: GSM92 form: Illumin t descriptor: forward eriment attri EO Accessio al: 4 runs, 25	butes: a: GSM927238 2.4M spots, 9.10	G bases, <u>5.6Gb</u>	2	
sam Libra Platf Spot 1 Expe GE Fota	ary: GSM92: form: Illumin t descriptor: forward eriment attri EO Accessio al: 4 runs, 25:	Itel Itel (more) ia (more) ibutes: nr. GSM927238 2.4M spots, 9.10 # of Spots # of Spots	G bases, <u>5.6Gb</u>	Size	
Libra Libra Platf Spot 1 Expe <i>GE</i> Fota	ary: GSM92 form: Illumin t descriptor: forward eriment attri EO Accessio al: 4 runs, 25 Run SRR494099	Butes:	G bases, <u>5.6Gb</u> (# of Bases 510M	Size 352.6Mb	
Libra Platf Spot Spot Expe GE Fota # 1. §	ary: GSM92 form: Illumin t descriptor: forward eriment attri EO Accessio al: 4 runs, 25 Run SRR494099 SRR494100	Butes: m: GSM927238 2.4M spots 9.10 # of Spots 14,166,619 104,643,824 104,643,824	G bases, <u>5.6Gb</u> (# of Bases 510M 3.8G	Size 352.6Mb 2.3Gb	
aibra libra Platf Platf 1 1 Expect GE GE Tota # 1. § 2. § 3. §	ary: GSM92 form: Illumin t descriptor: forward eriment attri EO Accessio al: 4 runs, 25 Run SRR494099 SRR494100 SRR494101	2238: HeLa (more) butes: m: GSM927238 2.4M spots, 9.10 # of Spots 14,166,619 104,643,824 105,956,875	G bases, <u>5.6Gb</u> (# of Bases 510M 3.8G 3.8G	Size 352.6Mb 2.3Gb 2.4Gb	
ibra latt Spot	ary: GSM92 form: Illumin t descriptor: forward	7238: HeLa (mo a (more)	v)		
n ora itf of 32	ary: GSM92: form: Illumin t descriptor: forward eriment attri EO Accessio al: 4 runs, 25:	2238: HeLa (more) butes: bn: GSM927238 2.4M spots, 9.10	G bases, <u>5.6Gb</u>	Size	
ibra libra ibra ipol ipol ispol i i i i i i i i i i i i i i i i i i i	ary: GSM92 form: Illumin t descriptor: forward eriment attri EO Accessio al: 4 runs, 25 Run SRR494099	butes: 	G bases, <u>5.6Gb</u> (# of Bases 510M	Size 352.6Mb	
sam ibra Platf Spot Spot 1 Expe GE GE Cota	ary: GSM92: form: Illumin t descriptor: forward eriment attri EO Accessic al: 4 runs, 25: Run SRR494099 SRR494100	2238: HeLa (more) butes: m: GSM927238 2.4M spots, 9.10 # of Spots 14,166,619 104,643,824	G bases, <u>5.6Gb</u> (# of Bases 510M 3.8G	Size 352.6Mb 2.3Gb	
Libra Platt Platt Spot 1 Expe GE GE Cota 4 1. § 2. § 3. §	Run SRR494100 SRR494100 SRR494100 SRR494100	2238: HeLa (more) butes: br: GSM927238 2.4M spots, 9.10 # of Spots 14,166,619 104,643,824 105,956,875	G bases, <u>5.6Gb</u> (# of Bases 510M 3.8G 3.8G	Size 352.6Mb 2.3Gb 2.4Gb	
# 1. 5 7 ota 1 5 pol 1 1 5 pol 1 1 5 pol 1 7 ota 1. 5 7 ota 3. 5 4. 5	ary: GSM92 form: Illumin t descriptor: forward eriment attri EO Accessic al: 4 runs, 25: Run SRR494099 SRR494100 SRR494101 SRR494102	# of Spots # of Spots 14,166,619 106,638,624 105,956,875 27,626,583	G bases, <u>5.6Gb</u> # of Bases 510M 3.8G 994.6M	Size <u>352.6Mb</u> <u>2.3Gb</u> <u>2.4Gb</u> <u>591.6Mb</u>	

Now go to the EBI SRA page using the following URL substituting "SRR_number" for yours:

http://www.ebi.ac.uk/ena/data/view/SRR_number

eg: http://www.ebi.ac.uk/ena/data/view/SRR494099

You will be taken to the ENA – European Nucleotide Archive.

Search		
Search		
Search		
Search		
Gealch		
E		
lead: XMI		
Feedback		
ise Count		
nload: <u>TEXT</u>		
(ftp) files		
(galaxy)		

Look for the fastq files (ftp) link and right-click on the link. A pop-up menu will appear – select Copy Link:

Navigation	Read Files											
This table con	tains the files f	or run SRR494	099									
Download file:	8											
View: TEXT										Download:	TEXT	
Select column	<u>s</u>											
Showing resu	lts 1 - 1 of 1 re	esults										
Study	Secondary	Sample	Experiment	Run	Scientific	Instrument	Library	Fasto	Fasto	Submitted	Submitted	
accession	study accession	accessions	accession	accession	name	model	layout	files (ftp)	files (galaxy)	files (ftp)	files (galaxy)	
SRP012667	SRP012667	SRS312185	SRX145994	SRR494099	Homo	Illumina	SINGLE	File	Open L	ink in New	Window	
					apiena	Analyzer II			Open L	ink in New	Tab	
									Downlo	oad Linked	File	
For Aspera do	or Aspera download, please download and install Aspera Connect								Downlo	oad Linked	File As	
							_	Add Li	nk to Book	marks		
									Add Li	nk to Read	ing List	
IBL-EBI		Ser	vices	Sale Sale	Res	search			Copy L	.ink		ĺ
ws		By	topic		Ove	erview			Add to	iTunes as	a Spoken Track	
ochures		By	name (A-Z)		Pub	lications			Train	at EBI		r

Paste this link into a text file, eg:

ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR494/SRR494099/SRR494099.fastq.gz

You can use this link with the unix command 'wget' to download the fastq file; connect to your CBRG account and move to your HTS space – *do not download HTS data under your home directory!* (please contact CBRG if you do not know where your HTS space is)

Then type

wget ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR494/SRR494099/SRR494099.fastq.gz &